

## Fieldwork phase 2

## Sample culture-based testing

Standard Operating Procedure

v2

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Project title: Feasibility study with a non-blinded, pilot, randomised baseline trial to evaluate the fidelity of a play space intervention model to reduce infant *Campylobacter* infection in rural households in Sidama Zone, Ethiopia

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This SOP is adapted from the CAGED study SOP with approval from the CAGED study Primary Investigators.

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## 1. Campylobacter testing: Safety considerations<sup>1</sup>

- Campylobacter species are Hazard Group 2 organisms and their infectious dose is 500 organisms by ingestion. The most effective method for preventing laboratoryacquired infections is the adoption of safe working practices
- Appropriate personal protective equipment and techniques designed to minimise exposure of the laboratory workers should be worn and adhered to at all times
- This includes a laboratory coat, goggles, gloves
- Good occupational hygiene practices should be followed, especially washing with warm water and soap
- All use of C. jejuni should be undertaken within the CAT 2 microbiology hood
- This protocol should be supplemented with approved COSHH and risk assessments.

 $https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\_data/file/685065/ID\_23i3.1.pdf$ 

## 1. Media preparation

#### 1.1 Preparation of blood-free broth for sample enrichment

Broth medium: Blood Free Campylobacter Broth

Selective supplement: Thermo Scientific™ Oxoid™ CCDA Selective Supplement Directions:

- 1. Suspend 16.75 grams in 500 mL distilled water
- 2. Heat to dissolve the medium completely
- 3. Sterilize by autoclaving at 15 lbs. pressure (121°C) for 15 minutes
- 4. Cool to 45-50°C
- 5. Aseptically add 1 vial of CCDA-Selective-Supplement
- 6. Mix well before dispensing into sterile containers

#### 1.2 Preparation of CHROMagar plates – calculation for 1 L

Campylobacter growth is on CHROMagar<sup>™</sup> at 42°C for 40–48 hrs. Campylobacter growth is on CHROMagar<sup>™</sup>. Preliminary identification of Campylobacter species from primary culture is by colonial appearance. On CHROMagar<sup>™</sup>, colonies appear as intense red coloured colonies on a translucent agar.

<sup>&</sup>lt;sup>1.</sup> Public Health England. (2018). Identification of Campylobacter species. UK Standards for Microbiology Investigations. ID 23 Issue 3.1.

#### COMPOSITION

The product is compos	ne product is composed of a powder base (B) and 1 supplement (S).				
Product =	Base (B)	Supplement (S)			
Total g/L	51.2 g/L	0.21 g/L			
Composition g/L	Agar 15.0 Peptone and yeast extract 25.0 Salts 9.0 Chromogenic and selective mix 2.2	Chromogenic and selective mix 0.21			
Aspect	Powder Form	Powder Form			
STORAGE	15/30 °C	2/8 °C			
FINAL MEDIA pH	7.4 +	-/- 0.2			

1	Final Media	HELPING CALCULATION
	1 L	0.21 g into 10 ml of purified water
	5 L	1.05 g into 50 ml of purified water

## Step 1: Preparation of the base

- Disperse slowly 51.2 g of powder base in 1L of purified water
- Stir until agar is well thickened
- Heat and bring to high temp (90-95°C) while swirling or stirring regularly

## DO NOT HEAT TO MORE THAN 95 °C. DO NOT AUTOCLAVE AT 121°C

• Cool in a water bath to 45-50 °C. Swirl or stir gently to homogenize

### Step 2: Preparation of the supplement

- In a transparent vessel, add 210 mg of supplement in 10 ml of purified water
- Swirl well until complete dissolution
- Filter to sterilize at 0.45 μm

## Step 3: Base + supplement

- Add the 10 ml of the supplement solution to the melted base (Step1) when the base has reached 45-50 °C
- Swirl or stir gently to homogenize

#### Step 4: Pouring

- Pour into sterile Petri dishes
- Let it solidify and dry

#### Step 5: Storage

- Store in the dark before use
- Prepared media plates can be kept for one day at room temperature
- Plates can be stored for up to 1 month under refrigeration (2/8 °C) if properly prepared and protected from light and dehydration
- Note: If not fully used, rehydrated CHROMagar Campylobacter supplement can be stored one month at 2-8°C or at -20°C

If the agar plate has been refrigerated, allow to warm to room temperature before inoculation.

#### 2. Plating samples

Note: all plating should take place under a laminar flow cabinet.

#### 2.1 Direct plating of faecal samples

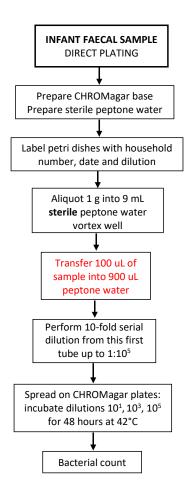
- 1. Label CHROMagar petri dishes with the household number, the date of the incubation and the dilution series
- 2. Collect 1 g of infant faeces. Record the exact faecal weight collected
- 3. Remove the air from the bag and return the bag to store the bag at 4°C until needed
- 4. Transfer the 1 g of faecal sample into a 15 mL sterile tube containing 9 mL of peptone water and vortex well with a vortex mixer
- Take 100 uL of sample using pipette into a sterile centrifuge tube containing 900 uL of peptone water and perform 10-fold serial solution up to 1:10<sup>5</sup> dilution
- 6. Vortex very well each tube before transferring the 100 uL into a new tube. Change pipette tips between dilutions
- 7. Take 100 uL of the diluted samples and spread the dilution series as below on **pre- plated/pre-labelled** CHROMagar petri dishes (10<sup>1</sup>,10<sup>3</sup>,10<sup>5</sup>)
- 8. Do not change the pipette tip if you go from a higher to a lower dilution but you must change tips between samples
- Incubate at 42°C for 48 hours in microaerophilic conditions. This is done by placing
  the stack of petri dishes into the microaerophilic jar with 1 sachet of CampyGen per
  stack of 10 petri dishes in each 2.5 L jar
  - (e.g. in a 2.5 litre jar, 1 sachet is required, in a 7.5 L jar, 3 sachets are required).
- 10. As soon as the sachets are opened, the contents activate, so place the sachets in the jar and seal the jar quickly
- 11. **Use the log sheet** to note the time the plate is cultured, infant ID number and the time the plate must come out

Note: all reading of colonies should take place under a laminar flow cabinet. Be careful when removing the lid of the jar and taking stacks of plates out – high contamination risk!

12. After 48 hours, count the colonies on each dilution plate and **record the data on the log sheet** 

Commented [SB1]: Note: I can't remember if we did this step or if we just dispensed 100 uL of the vortexed sample and did a serial from that – I expect so as that would've been the first concentration. This is highlighted in red on the diagram below, so see if you need to skip this step

Figure 1. Testing flowchart for Campylobacter isolation, growth and enumeration



Whirl-Pak™ sterile bag with scoop	https://www.sigmaaldrich.com/catalog/product/sigma/wpb01478wa?lang=en&region=GB
Buffered dehydrated peptone water	https://www.fishersci.co.uk/shop/products/oxoid-buffered-peptone-water/p-4524834#?keyword=buffered+peptone+water
CHROMagar™ powder	Quoted from BioConnections: https://www.bioconnections.net/chromagar.html
CHROMagar supplement	Quoted from BioConnections: <a href="https://www.bioconnections.net/chromagar.html">https://www.bioconnections.net/chromagar.html</a> . Chromagar website for US customers is: <a href="https://drg-international.com/products/chromagar/">https://drg-international.com/products/chromagar/</a>
0.33 um filters, syringe	From university
Blood Free Campy Broth 500g	https://www.sigmaaldrich.com/catalog/product/sial/59751?lang=en&region=GB
CCDA Selective Supplement	https://www.sigmaaldrich.com/catalog/product/sial/77093?lang=en&region=GB
Oxoid™ CampyGen™ 2.5L sachet (packs of 10)	https://www.fishersci.co.uk/shop/products/oxoid-campygen-2-5l-sachet/10108012#?keyword=campygen
Diamond shaped weigh boats	https://www.sigmaaldrich.com/catalog/product/sigma/hs1424aa?lang=en&region=GB
Petri dishes (90 mm)	From university
15 ml polypropylene centrifuge tubes (flat top cap, conical bottom)	From university
Microaerophilic jar (2.5 L, 7.5 L)	From university
Pocket digital weighing scale	https://tinyurl.com/y8np6jks
500 mL glass jar for prepared broth storage	From university
1000ul pipette	From university
200ul pipette	From university
L-Shaped spreaders	https://tinyurl.com/y3onc2wh
Incubator capable of 42°C	Lab
Laminar flow cabinet	Lab
Vortex mixer	Lab
Hot plate for agar prep	Lab
Nitrile gloves	Lab