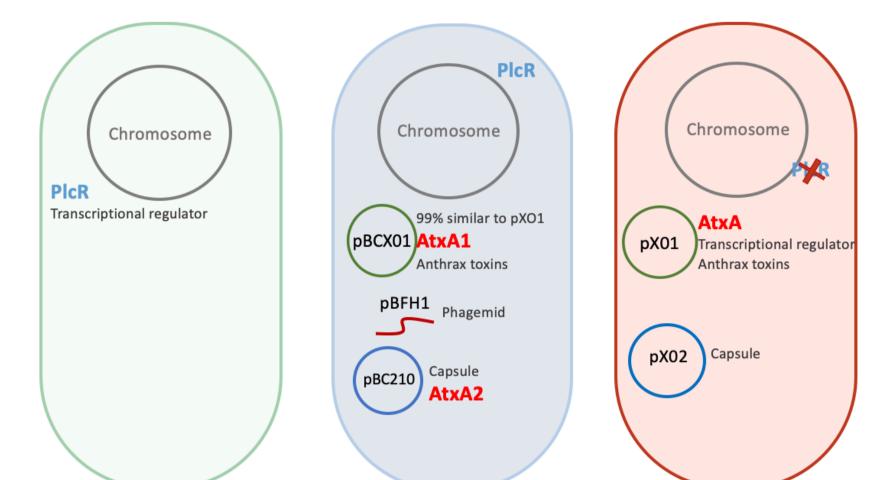
# Temperature dependent toxin expression in B. cereus G9241, the causative agent of anthrax-like illness

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## Background

- The bacteria *Bacillus cereus* G9241 is closely related to the anthrax agent *B. anthracis*.
- Members of the *B. cereus* group express **PlcR**, a transcriptional regulator of secreted toxins and proteases, which is activated by the peptide PapR.
- However, in all *B. anthracis* isolates, the *plcR* gene is truncated.
- It has been proposed that in *B. anthracis*, the acquisition of





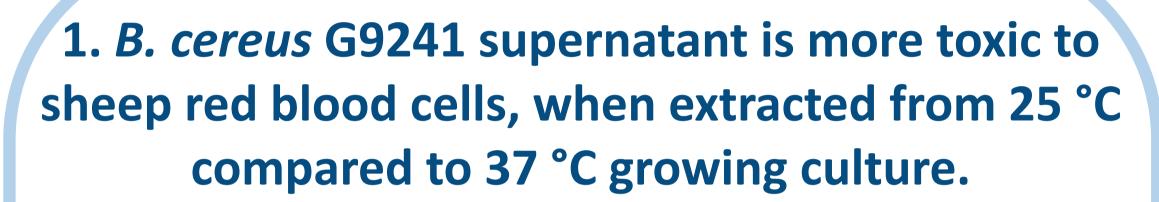
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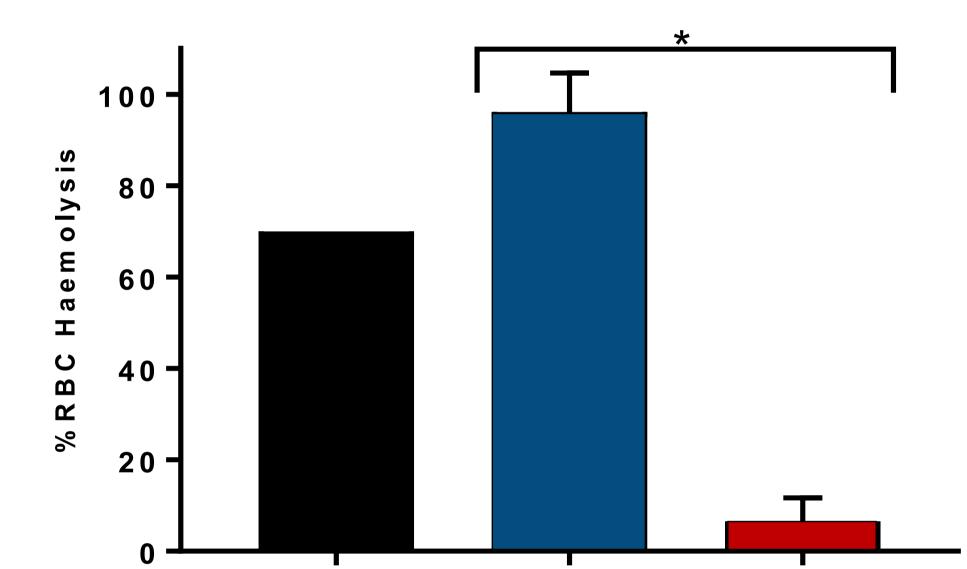
#### Aim

Investigate whether the activity of the PlcR-PapR regulatory PlcRcircuit and

AtxA, which regulates anthrax toxins, was incompatible with the activity of PlcR, leading to PlcR inactivation, and consequently restriction to mammalian hosts. Interestingly, G9241 encodes intact copies of both *atxA* and *plcR*.

• We suggest that, at least partial PlcR suppression is essential to facilitate the anthrax-like disease state in humans.





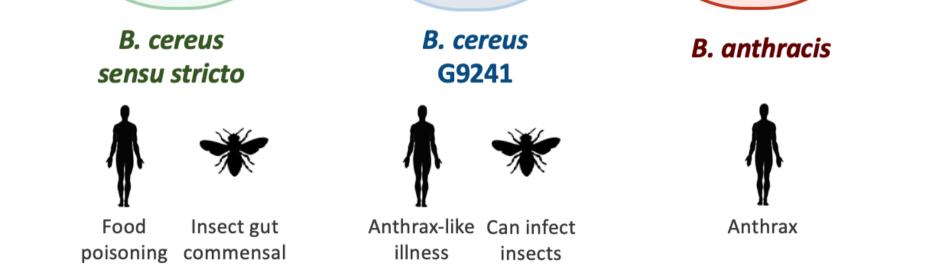
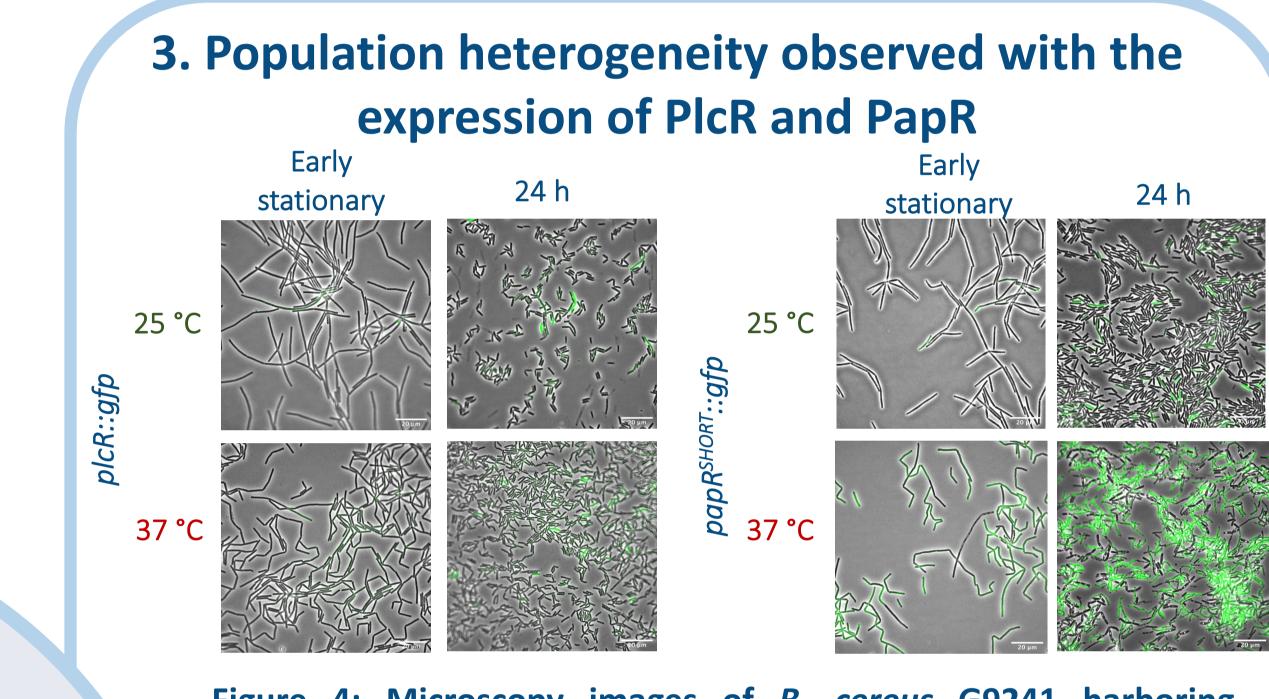


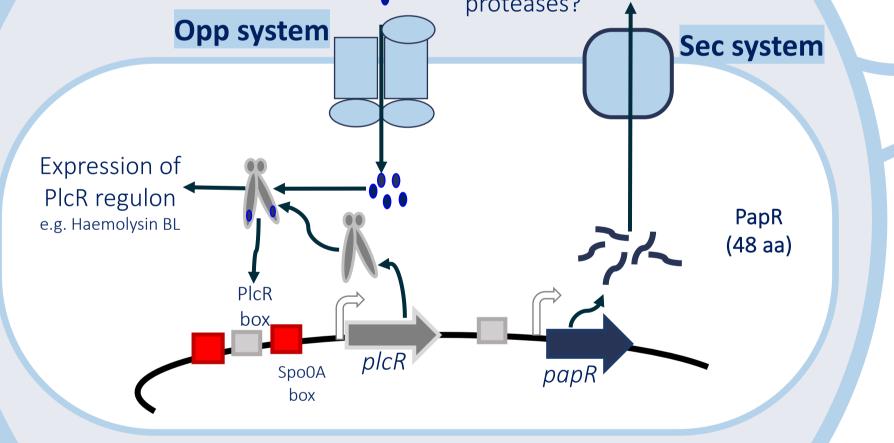
Figure 1: The Bacillus cereus group consists of many species which includes Bacillus cereus sensu stricto and Bacillus anthracis.

regulated toxins in B. G9241 cereus are temperature dependent, in order to accommodate the activity of AtxA.

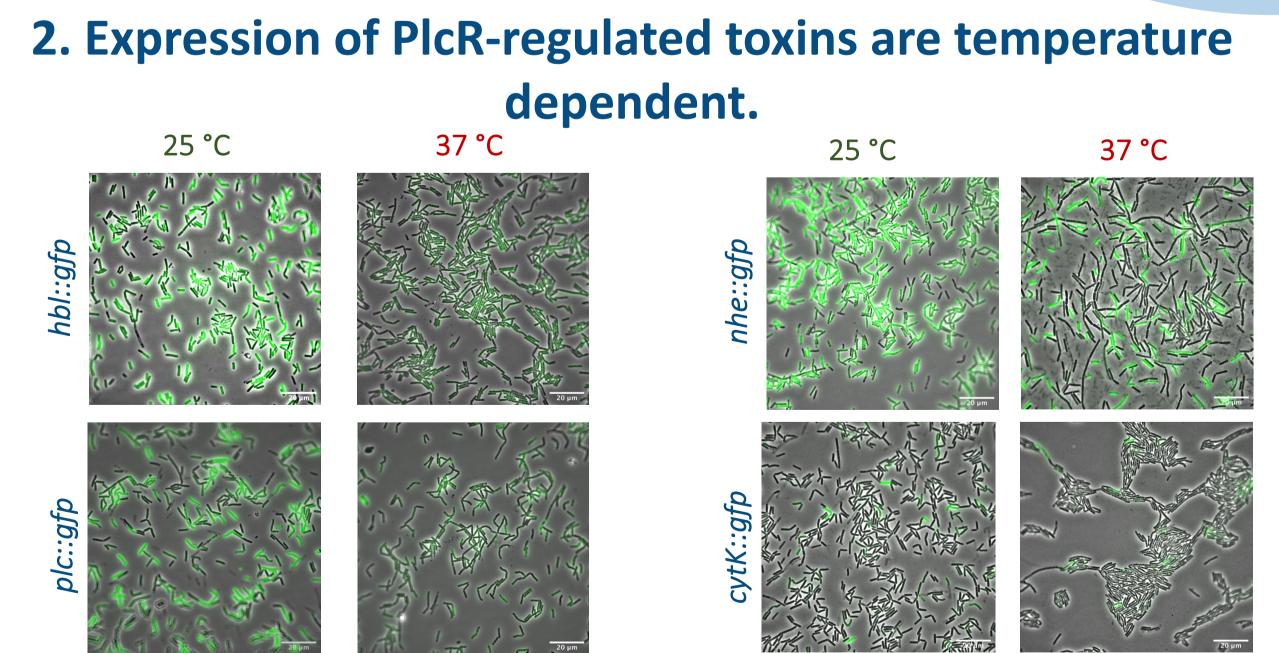


Microscopy images of *B. cereus* G9241 harboring Figure 4: papR<sup>SHORT</sup>::gfp. Plasmid-based plcR::gfp GFP or reporter

Figure 2: Haemolysis assay using *B. cereus* G9241 supernatant. B. cereus G9241 supernatant is significantly more toxic to sheep red blood cells, when extracted from 25 °C growing culture compared to 37 °C growing culture (Brooker et al., unpublished data). \* denotes an unpaired t-test with a p-value of 0.0232. Error bars denote one standard deviation and all samples were to an n=3.



**The PlcR-PapR** regulatory circuit



constructs.

4. Oligopeptide permease (Opp) system, which is involved in importing active PapR peptide, is functional at 37 °C

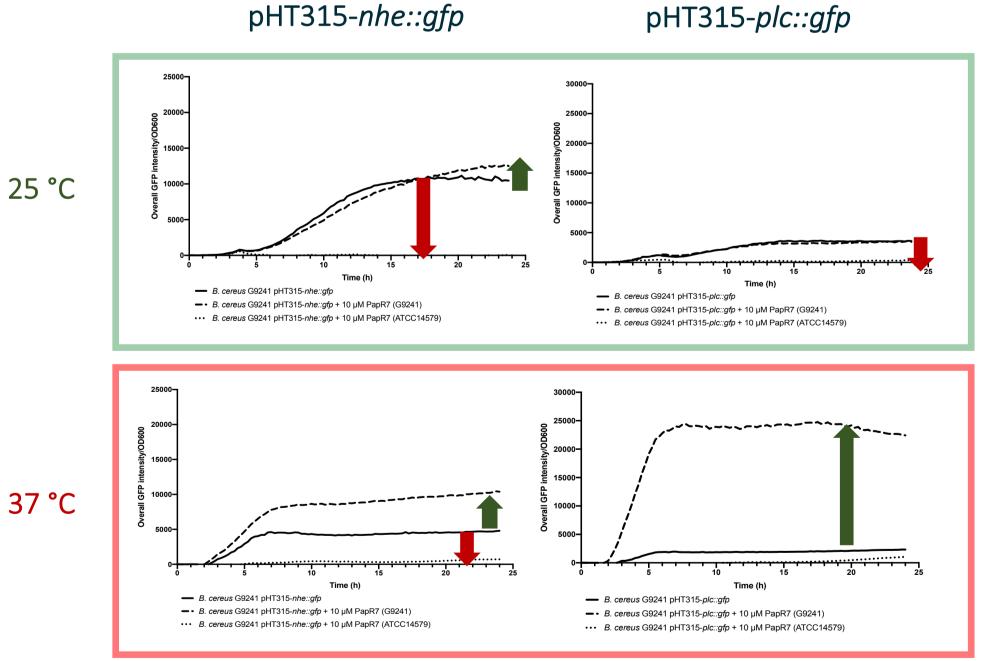


Figure 5: Overall GFP intensity per cell of *B. cereus* G9241 containing plasmid-based toxin reporters over time grown at 25 °C and 37 °C in LB media, with or without the addition of synthetic PapR<sub>7</sub>. N=3

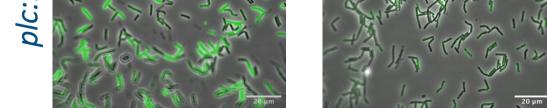


Figure 3: Microscopy images of *B. cereus* G9241 at 24 hours harboring plasmid-based GFP reporter constructs of PlcR-regulated toxins.

### REFERENCE

Hoffmaster, A. R., et al. (2004). Proceedings of the National Academy of Sciences of the United States of America **101**(22): 8449-8454.

Mignot, T. et al. (2001). <u>Molecular Microbiology</u>, 42(5), pp. 1189–1198.

Slamti, L. and Lereclus, D. (2005). Journal of Bacteriology, 187(3), pp. 1182–1187.

## Conclusion

- The expression of PlcR-regulated **toxins are higher at 25 °C** compared to 37 °C, which indicates why we see haemolytic activity at 25 °C.
- The export of unprocessed PapR (data not shown) and import of processed **PapR** is **functional at both temperatures**, suggesting that they do not play a role in the temperature dependent expression of toxins.
- This suggests that the protease(s) needed in processing PapR is possibly involved in the temperature dependent toxin expression.
- Similar results is observed in *B. cereus* G9241 lacking the pBCX01 plasmid (data not shown), which suggests that the AtxA does not modulate the activity of the PlcR-regulated toxins.